CUPARANE- AND ISOCUPARANE-TYPE SESQUITERPENOIDS IN LIVERWORTS OF THE GENUS HERBERTUS*

YOSHINORI ASAKAWA[†], REIKO MATSUDA[†], W. B. SCHOFIELD[‡] and S. ROB GRADSTEIN§

†Institute of Pharmacognosy, Tokushima Bunri University, Yamashiro-cho, 770 Tokushima, Japan; ‡Department of Botany, The University of British Columbia, 2075 Westbrook Mall, Vancouver, B. C., Canada; §Institute of Systematic Botany, Heidelberglaan 2, Utrecht, The Netherlands

(Received 9 February 1982)

Key Word Index—Herbertus aduncus; H. sakuraii; H. subdentatus; Jungermanniales; Hepaticae; cuparene; 2-hydroxycuparene; 3-hydroxycuparene; 2,3-dihydroxycuparene; herbertene; 2-hydroxyisocuparene; α -cuparenone; ent-sesquiterpenoids; chemosystematics.

Abstract—Canadian and Japanese Herbertus aduncus, Japanese H. sakuraii and Colombian H. subdentatus belonging to the Jungermanniales (Hepaticae) were chemically investigated. The previously known entcuparane- and novel isocuparane-type sesquiterpenoids are the major components which are of importance as chemical markers in the Herbertaceae.

INTRODUCTION

Most liverworts have oil bodies which contain mainly mono-, sesqui- and diterpenoids and/or lipophilic aromatic compounds. These substances are present as major components and thus we can apply them to a chemosystematic study of the Hepaticae [1-13].

Herbertus species belonging to the suborder Herbertineae are rather primitive liverworts in the Jungermanniales. In previous papers [2, 5, 8, 9], we noted that H. aduncus (Dicks.) S. Gray, H. sakuraii (Warnst.) Hatt. and H. subdentatus (Steph.) Fulf. produced cuparane- and isocuparane-type sesquiterpenoids. More recently, Matsuo et al. [14] reported the isolation of herbertene (isocuparene) (6) from H. aduncus. The present communication is concerned with the distribution of ent-cuparane- and ent-isocuparane-type sesquiterpenoids in three Herbertus

species, H. aduncus, H. sakuraii and H. subdentatus, collected in Canada, Colombia and Japan.

RESULTS AND DISCUSSION

Air-dried and ground material of each species was extracted with diethyl ether. The extracts were checked by TLC and GC and further analysed by GC/MS equipped with a computer. Chemical components were identified by GC co-injection of standard compounds, analysis of each mass spectrum and direct comparison of mass spectra with those of authentic samples. The major components were further isolated by prep. TLC and GC, and their structures confirmed by spectral evidence and chemical degradations. The species, sampling locations, date of collection and compounds detected in each *Herbertus* species were listed in Table 1.

The major components of *H. aduncus* (sample 1) are new isocuparane-type sesquiterpene alcohols, 2-hydroxyisocuparene [m-(1, 2, 2-trimethylcyclopentyl)-o-cresol] (8), 4-hydroxyisocuparene [m-(1, 2, 2-trimethylcyclopentyl)-o-cresol]

Table 1. Cuparane- and isocuparane-type sesquiterpenoids of Herbertus species

Sample	Species	Site and date				Compounds detected	detected				
<u>e</u>		or collection –	1	7	m	4	5	9	7	œ	Sterols‡
	Herbertus aduncus	Herbertus aduncus Mt Fujikura, Imajo-cho, Nanjo-gun, Fukui, Japan Oct. 1978	* +	+	l	+ + + + +	1	+ +	+ + + + +	+++	+
7	H. aduncus	Komagadake, Hakone, Kanagawa, Japan Aug. 1980	+	+	1	ļ	I	+++	+ + + +	+	+
8	H. aduncus†	Bridal Veil Falls, Popkum, Fraser River Valley, British Columbia, Canada May 1981	+	+	1	++	+	++	+ + +	+ + +	+
4	H. sakuraii	Yaku Island, Kagoshima, Japan Dec. 1979	I	+	1	l	ı	+	+ + + +	1	+
S	H. subdentatus	Paramo de Guasca, Columbia Aug. 1980	+ :	+	+	+ + + + + +	[+	+ + + + + + + + + + + + + + + + + + + +	+ +	+

*The symbols, +, + +, + + + etc are the relative concentrations estimated by GC/MS. $\dagger \beta$ -Pinene and 1-octen-3-ol were detected in this species as minor components. \dagger Campesterol, stigmasterol and sitosterol.

trimethycyclopentyl)-p-cresol] (7) and a new sesquiterpene diol, 2, 3-dihydroxycuparene (4) in which 7 is the most abundant. Cuparene (1), herbertene (isocuparene) (6) and 2-hydroxycuparene (2) were also detected as minor components. H. aduncus was further collected in two different localities, Kanagawa (Japan) (sample 2) and British Columbia (Canada) (sample 3). The gas chromatogram of the crude extract (sample 2) was simpler than that of sample 1. The major component of sample 2 was 4-hydroxyisocuparene (7). However, 2, 3-dihydroxycuparene (4) was not detected in sample 2, even in GC/MS analysis. It is not clear from the present results whether the presence or absence of 2, 3-dihydroxycuparene in the two Japanese H. aduncus is caused by seasonal or geographical variation. The chemical constitution of Canadian H. aduncus is quite similar to that of the same species in Japan (sample 1), except for the presence of an additional cuparane-type sesquiterpene ketone, α -cuparenone (5), as a minor component.

H. sakuraii synthesized 4-hydroxyisocuparene (7) and 2-hydroxyisocuparene (8), together with herbetene (6). Thus, H. sakuraii is chemically quite close to H. aduncus. Colombian H. subdentatus also produced the same cuparane- and isocuparane-type sesquiterpenoids as H. aduncus and H. sakuraii and their chemical profiles quite resemble that of H. aduncus (sample 1). In addition to the above components, H. subdentatus contains 3-hydroxycuparene (3) whose structure was tentatively assigned by GC/MS analysis.

In previous communications [2, 5, 8, 9], we reported that 2-hydroxycuparene (2) was the major component of *Herbertus* species. Further chemical and spectral data of each of the isolated cuparene- and isocuparene-type sesquiterpenes showed that the major constituents were hydroxyisocuparenes (7, 8) whose mass spectra were quite similar to those of 2-hydroxycuparene (2).

From the above results, it is clear that isocuparaneand cuparane-type sesquiterpenoids are characteristic constituent of *Herbertus* species. Recently, we reported that the chemical markers of *Trichocolea* tomentella, which is referred to the suborder Herbertineae in the modern classification of the Hepaticae [15], are isoprenyl benzoates [13]. It is apparent therefore, that there is no chemical affinity between Herbertus and Trichocolea species. The present cuparane- and isocuparane-type sesquiterpenoids belong to the ent-series and are thus characteristically distinctive in stereochemistry from the sesquiterpenoids of higher plants.

EXPERIMENTAL

GC/MS spectra were obtained under the following conditions: electrical energy 70 eV; trap current 60 μ A; temp. 270°. GC column: 5% SE-30, 3m × 2 mm glass column: temp. 50-270° at 5°/min, inject temp. 260°, He 30 ml/min. Analytical and prep. GC: 5% SE-30, 3m × 2 mm and 5% DEGS glass column, temp. programme 50-270° at 5°/min, inject temp. 260°, N₂ 30 ml/min. TLC and prep. TLC: pre-coated Si gel (0.25 mesh) F₂₅₄, n-hexane-EtOAc or C₆H₆-EtOAc (4:1) as solvents. Compounds were detected by I₂ vapor, 30% H₂SO₄ or UV light (254 nm).

Plant materials. Herbertus species identified by W. B. S.,

S. R. G., Drs. S. Hattori and H. Inoue were deposited in the Herbaria of Institute of Pharmacognosy, Tokushima Bunri University, Universities of Bogota and Utrecht.

Extraction and isolation. Each Herbertus species sample was air-dried for 5 days, ground mechanically and the powder (250 mg) extracted with Et₂O for 5 days. The extracts were filtered through a short glass column packed with Si gel (230-400 mesh) and the filtrates directly analysed by TLC, GC and GC/MS equipped with a computer, after removal of the solvent. The remaining materials were also extracted with Et₂O for 1 month and then re-extracted with MeOH for 2 weeks. Each extract was combined and the crude oils were chromatographed on Si gel using a n-hexane-EtOAc or C₆H₆-EtOAc gradient. The chemical structures of the isolated compounds by prep. TLC and GC were confirmed by IR, 1H NMR, and MS analyses and some chemical degradations. The structure determination of the new isocuparenes and 2,3-dihydroxycuparene from Herbertus species will be reported elsewhere.

Acknowledgements—We are indebted to Drs. S. Hattori and M. Mizutani and Z. Iwatsuki (Hattori Botanical Laboratory, Nichinan, Japan) and H. Inoue (National Science Museum of Tokyo) for their collection and identification of Japanese Herbertus species and for useful suggestions.

REFERENCES

- Asakawa, Y., Tokunaga, N., Toyota, M., Takemoto, T. and Suire, C. (1979) J. Hattori Bot. Lab. 45, 395.
- Asakawa, Y., Tokunaga, N., Toyota, M., Takemoto, T., Hattori, S., Mizutani, M. and Suire, C. (1979) J. Hattori Bot. Lab. 46, 67.
- Asakawa, Y., Hattori, S., Mizutani, M., Tokunaga, N. and Takemoto, T. (1979) J. Hattori Bot. Lab. 46, 77.
- Asakawa, Y., Tokunaga, N., Takemoto, T., Hattori S., Mizutani, M. and Suire, C. (1980) J. Hattori Bot. Lab. 47, 153.
- Asakawa, Y., Suire, C., Toyota, M., Tokunaga, N., Takemoto, T., Hattori, S. and Mizutani, M. (1980) J. Hattori Bot. Lab. 48, 285.
- Asakawa, Y., Inoue, H., Toyota, M. and Takemoto, T. (1980) Phytochemistry 19, 2623.
- Asakawa, Y., Matsuda, R., Takemoto, T., Hattori, S., Mizutani, M., Inoue, H., Suire, C. and Huneck, S. (1981) J. Hattori Bot. Lab. 50, 107.
- Asakawa, Y., Matsuda, R., Toyota, M., Suire, C., Takemoto, T., Inoue, H., Hattori, S. and Mizutani, M (1981) J. Hattori Bot. Lab. 50, 165.
- Gradstein, S. R., Matsuda, R. and Asakawa, Y. (1981) J. Hattori Bot. Lab. 50, 231.
- Asakawa, Y., Matsuda, R. and Suire, C. (1981) Phytochemistry 20, 1427.
- Asakawa, Y., Matsuda, R., Toyota, M., Hattori, S. and Ourisson, G. (1981). Phytochemistry 20, 2187.
- Toyota, M., Asakawa, Y. and Takemoto, T. (1981) Phytochemistry 20, 2359.
- 13. Asakawa, Y., Toyota, M., Takemoto, T. and Mues, R. (1981) Phytochemistry 20, 2695.
- 14. Matsuo, A., Yuki, S., Nakayama, M. and Hayashi, S. (1981) J. Chem. Soc. Chem. Commun. 864.
- Schuster, R. M. (1979) In Bryophyte Systematics (Clarke, G. C. S. and Duckett, J. G., eds.) pp. 41-82. Academic Press, New York.